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Active Noise Experienced by a Passive Particle Trapped in Active Bath

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INTRODUCTION

Active colloids consist of mesoscale self-propelled units that are able to locally convert stored or ambient energy into their persistent locomotion [1–3]. Prominent examples include colonies of bacteria [4–6] and synthetic microswimmers [7–13]. Due to being intrinsically out of equilibrium, active colloidal suspensions can serve as an active bath that can profoundly impact the structure and dynamics of immersed passive objects. For instance, active bath can greatly deform flexible macromolecules [14–19], give rise to unusual effective interactions between passive objects [20–22] or unexpected self-assemblies [23–27]. Moreover, active bath can dramatically enhance the diffusion of passive tracers by orders of magnitude compared to thermal cases [28, 29], and can drive a directed motion of asymmetric objects [6, 30–32]. These phenomena are fundamentally different from equilibrium thermal baths. Despite their important differences, the generalization of important physical concepts and relations (such as temperature, pressure, depletion force, equation of state or fluctuation relation) from thermal to active is completely determined by the active swimmer, the passive tracers and their interactions [43–49], which constitutes a paradigm to study nonequilibrium statistical physics [50–52]. The active noise, although artificially imposed in theories [46–49], can be directly extracted from colloidal experiments or simulations. It is implicitly assumed that the active noise is an intrinsic feature of the active bath, similarly to the thermal noise in equilibrium, and is completely determined by the active swimmers, the passive tracers and their interactions [43–49]. Recent studies, however, suggest that active pressure depends on the details of external boundary walls [33] and active deformation is drastically changed with external constraints [54]. These observations raise the fundamental question whether and how external conditions influence the active noise.

In this paper, we study the dynamics of a passive particle suspended in an active bath, trapped by harmonic potentials of various stiffness, using computer simulations and theoretical analyses. In simulations, the active bath is modelled by an ensemble of active Brownian particles (ABP); and in theory, the active bath is described by an active noise. By fitting the theoretical prediction to the simulation measurement, we find that the extracted active noise, both its magnitude and correlation time, sensitively depends on the constraint experienced by the passive particle, in contrast to its thermal counterpart. An intuitive explanation for this constraint dependence of active noise is that the external trapping increases the intensity of collisions between the tracer and active particles. Theoretical model with a constraint-dependent active noise quantitatively reproduces the mean potential...
energy of the tracer particle measured in simulations. Furthermore, we show that three widely used effective temperatures (separately quantified as the ratio of fluctuation to dissipation, the average potential and kinetic energies of the passive particle) all change significantly with the external constraint, but in different manners. Our results demonstrate that the active noise experienced by a confined passive tracer cannot be treated as an intrinsic physical quantity of the active bath, and a single constant effective temperature is not sufficient to describe the tracer behaviors.

**SIMULATION AND THEORY**

*Simulation.*—The simulation is performed by considering a 2D system consisting of 2000 small ABPs with diameter of $\sigma_s$ and mass of $m$, and one large passive tracer with diameter of $\sigma_l = 3\sigma_s$ and mass of $M$, as sketched in Fig. 1(a). The interactions between the particles are modelled by a repulsive Lennard-Jones-type potential, $U(r) = 4\epsilon \left( \left( \frac{\sigma_s}{r} \right)^{12} - \left( \frac{\sigma_s}{r} \right)^{6} \right) + \epsilon$, for $r \leq 2\pi \sigma_s$. The interaction diameter between passive and active particles is taken as $\frac{1}{2}(\sigma_s + \sigma_l)$. The translational motion of the active particles evolves according to the underdamped Langevin equation,

$$m\dot{\mathbf{v}} = -\gamma_s \mathbf{v} + \mathbf{F}_d + \mathbf{F}_r + \xi,$$

with $\mathbf{F}_d$ the self-propulsion force, $\mathbf{F}_r$ the steric interaction between particles, $\gamma_s$ the friction coefficient, and $\xi$ the Gaussian distributed stochastic force with zero mean and variance $\langle \xi_a(t)\xi_b(t') \rangle = 2k_bT\gamma_s\delta(\tau - \tau')$ ($\alpha$ and $\beta$ represent Cartesian components). In the simulations, we always take $F_d\sigma_s/k_bT = 20$, which is comparable to the value of the swimming E.coli bacteria in experiments. The Brownian rotation of the active particle is similarly simulated with the rotational friction coefficient $\gamma_r = \frac{1}{2}\sigma_s^2\gamma_s$.

The translational motion of the active particle also follows Eq. (1) with the frictional coefficient $\gamma_t = 3\gamma_s$ and $\mathbf{F}_d = 0$. Additionally, the passive tracer is constrained by a harmonic potential $U_{ext}(r) = \frac{1}{2}kr^2$, with $r$ the tracer displacement from the trap center and $k$ the trap stiffness. A typical trajectory of the large passive particle trapped in the active bath is plotted in Fig. 1(b).

In the simulations, we use the underdamped (instead of overdamped) Langevin equation in order to quantify the kinetic energy of the passive tracer.

*Theory.*—The effect of the active bath on the passive tracer can be theoretically captured by including an active colored noise term in the traditional overdamped Langevin equation [43–49],

$$\dot{\mathbf{r}} = \mu \mathbf{f}(\mathbf{r}) + \eta + \eta^A,$$

with $\mathbf{f}(\mathbf{r}) = -kr$ the restoring force from the external trap, $\mu$ the tracer mobility, $\eta$ the thermal noise and $\eta^A$ the active noise. The variance of the thermal noise reads $\langle \eta_a(t)\eta_b(t') \rangle = 2D_T\delta(\tau - \tau')$ with $D_T$ the diffusion coefficient of the passive tracer originating from the thermal bath. The variance of the active noise is $\langle \eta^A_a(t)\eta^A_b(t') \rangle = D_A\delta(\tau - \tau')$, since $D_A$ denotes the active contribution to the tracer diffusion coefficient characterizing the strength of the active fluctuations and $\tau$ is the correlation time of the active fluctuations. Similar coarse-grained model has been widely used to describe active colloidal particles themselves [55–60], where $\mathbf{f}(\mathbf{r})$ corresponds to the interactions between the active particles.

In previous work based on Eq. (2) [43, 45–49], the diffusion coefficient $D_T$ is taken as $D_T = \mu k_bT$, with the mobility $\mu$ only originating from the thermal bath. This prescription does not consider the active friction contributed by the couplings between the tracer and active particles [49], and is only valid in the dilute limit for active baths [50]. For semidilute or concentrated active particle solutions, the $\mu$ needs to include both the thermal and active contributions, so that the average friction is balanced by the external force, i.e. $\dot{\mathbf{r}}/\mu = (\mathbf{f}(\mathbf{r}))$ (the friction memory kernel remains instantaneous [44]). In these cases, the thermal diffusion coefficient $D_T$ does not obey the Einstein relation $D_T = \mu k_bT$, since $\mu$ now contains the active contributions. In simulations, the $D_T$ can be measured as the diffusion coefficient of the tracer in the same active bath when the activities of the constituent swimmers are switched off, such that $D_A$ exclusively corresponds to the active diffusion following its definition.

Following Ref. [43], the mean square displacement (MSD) of the passive tracer in the $x$ (or $y$) direction can be calculated from Eq. (2) as

$$\langle \Delta x^2(t) \rangle = \frac{2D_T}{\mu k}(1 - e^{-\mu k t})$$

$$+ \frac{2D_A}{\mu k} \frac{1 - e^{-\mu k t} - \mu k (1 - e^{-\mu k t})}{1 - (\mu k t)^2}.$$  

*Fig. 1: (a) Schematic diagram of a passive particle (grey) subjected to an external harmonic potential in an active bath consisting of ABPs (Jaanus). The self-propelling direction of the ABP is marked with the red cap. (b) The motion trajectory of the passive particle in the harmonic trap, which is represented by the colored area.*
Using Eq. (3) to fit the simulation MSD with the independently measured $D_T$ and $\mu$, the strength $D_A$ and the relaxation time $\tau$ of the active noise experienced by the passive tracer can be directly extracted.

RESULTS AND DISCUSSION

We first measure the MSD of the passive tracer particle in a semidilute active solution with the packing fraction of the microswimmers $\rho = 0.3$ for various trap stiffness $k$. Two representative MSD curves with $k\sigma^2/k_BT = 40$ and 120 are shown in Fig. 3(a). The MSD data can be well fitted to Eq. (3) using the $D_A$ and $\tau$ as fitting parameters, with the $D_T \simeq (0.47 \pm 0.02)k_BT/\gamma l$ and $\mu \simeq (0.59 \pm 0.01)/\gamma l$ that are independently measured in simulations. Here, $D_T$ is obtained from the MSD of a passive tracer freely moving in a solution of inactive microswimmers (without activity, $F_d = 0$), and $\mu$ is quantified by the drift velocity of a free passive tracer $v_d$ under an externally applied force $F_e$ in the active bath. To properly determine $\mu$, the applied force is so small that the system is in the linear response regime, as demonstrated by the linear relation between $v_d$ and $F_e$ in Fig. 2. Note that, in the semidilute situation, the measured $D_T$ and $\mu$ significantly deviate from the Einstein relation $D_T = \mu k_BT$, as analyzed in the theory above. For comparison, we also consider a dilute active solution with the microswimmer packing fraction $\rho = 0.06$. The MSD of the passive particle can still be well fitted to Eq. (3), as plotted in Fig. 3(c). In this case, the Einstein relation $D_T = k_BT\mu$ is maintained as expected, with $D_T \simeq (0.88 \pm 0.04)k_BT/\gamma l$ and $\mu \simeq (0.89 \pm 0.01)/\gamma l$ close to its thermal counterpart.

The quantities $D_A$ and $\tau$ obtained from the MSDs show clear dependence on the trap stiffness $k$ in both semi-dilute and dilute active baths, as plotted in Figs. 3(b) and (d). The strength of the active fluctuations monotonically increases with $k$ for both packing fractions, while the relaxation time of the active noise exhibits different dependence on $k$ for the semidilute and dilute active baths. The dependence of $D_A$ and $\tau$ on external constraint unambiguously indicates that the active fluctuations experienced by a passive tracer is not an intrinsic property of the active bath, and external constraints must be considered when active noise exists.

The increase of $D_A$ with $k$ can be understood from the collision kinetics between the tracer and the active swimmers. One of the distinctive differences between collisions with passive and active particles is that the momentum change in collisions with passive particles depends only on the momenta before the collision, while additional momentum can be transferred to the tracer from the persistent motion of the self-propelling swimmer. The intensity of the persistent collisions with microswimmers, thus $D_A$, can be greatly influenced by the constraints imposed on the passive tracer. Briefly, for strong constraints, most of the driving force $F_d$ is transferred to the tracer, leading to a large $D_A$; while for weak constraints, the friction on the colliding swimmer (moving with the tracer at a relatively large velocity) offsets part of $F_d$ and weakens the collision intensity. This picture can be qualitatively formulated by implementing an approximate calculation based on microscopic collision processes. For simplicity, we only consider a head-on collision between the passive and active particles (i.e., the swimmer orientation parallel to the vector connecting the swimmer and tracer). Further, we assume that before the collision, the passive

![FIG. 2: Reduced drift velocity $v_d$ of a free passive tracer as a function of externally applied force $F_e$ in both semi-dilute ($\rho = 0.06$) and dilute ($\rho = 0.06$) active baths. The dashed line denotes a linear fit to the measured data, with the slope corresponding to $\mu$.](image)

![FIG. 3: The MSD in $x$ direction for various external constraints $k\sigma^2/k_BT = 40$ (black open circles) and $k\sigma^2/k_BT = 120$ (blue open squares) at the packing fractions of the active particles $\rho = 0.3$ (a) and $\rho = 0.06$ (c). Solid lines are fits with Eq. (3). The dimensionless $D_A$ (black open circles) and $\tau$ (blue open squares) as functions of trap stiffness, extracted by fitting for $\rho = 0.3$ (b) and $\rho = 0.06$ (d).](image)
tracer rests at the trap center; and after the collision, the swimmer moves with the tracer for a duration of the orientational relaxation timescale of swimmer, \(\tau_r = \gamma_r/k_b T\). Disregarding the thermal stochastic force (only considering active collisions), the overdamped Langevin equation of the tracer particle reads, \((\gamma + \gamma_s)\dot{r} = -kr + F_d\), which is obtained by noticing that \(F_d = \gamma r\) corresponds to the force exerted on the tracer by the swimmer. By solving this equation, we obtain the tracer trajectory

\[
r(t) = \frac{F_d}{k}[1 - \exp(-kt/\gamma)],
\]

with \(\gamma = \gamma_l + \gamma_s\). The active stochastic force, \(\eta^A/\mu\), arising from the persistent collisions between the swimmer and tracer, is roughly proportional to \(F_d = \gamma r\). Hence, \(\langle \eta^A \cdot \eta^A \rangle / \mu^2 \sim \frac{1}{\tau} \int_0^\infty (F_d - \gamma_t \dot{r})^2 dt\), which can be evaluated using Eq. (4). With the equal-time correlation of the active noise \(\langle \eta^A(t) \cdot \eta^A(t) \rangle = 2D_A/\tau\), we obtain

\[
D_A \sim \frac{1}{2} \tau^2 \rho^2 F_d^2 [1 - \frac{2\gamma_t}{k}\{1 - \exp(-\tau/\gamma_t)\}] + \frac{\gamma_t^2}{2\tau} [1 - \exp(-2\tau/\gamma_t)],
\]

which monotonically increases with \(k\) and saturates at large \(k\), consistent with the simulation results in Fig. 3. These calculations do not consider the events of tracer simultaneously colliding with multiple swimmers or the swimmer-tracer collision frequency.

The \(k\) dependence of the relaxation time of the active noise can be understood by considering non-head-on collisions between the tracer and the swimmers. For non-head-on collisions in dilute solutions, strong constraints prevent the tracer particle from disengaging from the colliding swimmer, which results in a more persistent collision and hence a longer \(\tau\) (Figs. 3(d)), and vice versa. The decrease of \(\tau\) in semidilute solutions (Figs. 3(b)), however, is probably due to the accumulation of microswimmers around the passive tracer and the formation of multi-layer structure of swimmers [61, 62]. This accumulation effect enhances with increasing the constraint, as active swimmers are more likely to accumulate around an immobile tracer than around a mobile one. Surrounded by layers of active swimmers, the tracer experiences multiple collisions from all directions, decreasing the correlation time of the active noise.

Analogously to temperature that is related to the magnitude of thermal noise, effective temperature of active bath could reflect the strength of active noise to some extent. The concept of effective temperature provides a possible way to map nonequilibrium to equilibrium systems, and it has been often employed to describe the active systems based on the well-established equilibrium analyses [33–37, 43, 51]. So far, we have shown that the active noise is not an intrinsic property of the active bath and is significantly influenced by external potential fields. It is therefore reasonable to expect that the effective temperatures in active system might be constraint-dependent. In the following, we investigate how external constraints affect the effective temperature of the passive tracer in the active bath. We consider three widely-used definitions of effective temperature, which are separately quantified as the ratio of fluctuation to dissipation \((D_A + D_T)/\mu\) (red open diamonds, right longitudinal axis) as functions of the trap stiffness for \(\rho = 0.30\) (a) and \(\rho = 0.06\) (c). The effective temperatures defined by average potential energy \(U\) as a function of the trap stiffness for \(\rho = 0.30\) (b) and \(\rho = 0.06\) (d), where the diamonds and crosses correspond to the simulation measurement and theoretical calculation with Eq. (5). For comparison, the mean potential and kinetic energies of the tracer in an equilibrium bath are also provided (squares), in which the activities of the microswimmers are turned off.

Figure 4 plots the measured effective temperatures as a function of trap stiffness in different concentrations of active swimmers. The thermal temperatures in passive thermal baths are also plotted for comparison. All three effective temperatures depend on the stiffness of the external trap, although with different trends. The ratio of the fluctuation to dissipation \((D_A + D_T)/\mu\) is markedly higher than the thermal temperature, and increases with the trap stiffness until saturation at large \(k\). This trend can be qualitatively understood in terms of the \(k\) dependence of \(D_A\) discussed above, as \(D_T\) and \(\mu\) remain unchanged.

The second effective temperature \(E\) is obtained by directly measuring the average kinetic energy of the passive tracer, \(\frac{1}{2}M\langle V_0^2 + V_A^2\rangle\), in underdamped Brownian dynamics simulations. As plotted in Fig. 4(a) and (c), the \(E\) decreases with the trap stiffness, qualitatively opposite to \((D_A + D_T)/\mu\). The \(E\) dependence on \(k\), however, is relatively weak, with \(E\) only slightly higher than the thermal temperature. The explanation of this trend is straightforward. The tracer obtains additional kinetic energy from collisions with the microswimmers in the active bath. Consider a trapped passive tracer, its mean kinetic energy reads \(E = \frac{1}{2}M\langle V_0^2 + V_A^2\rangle = k_b T + \frac{1}{2}M\langle V_A^2\rangle\).
where $\mathbf{V}_0$ and $\mathbf{V}_A$ are the velocities contributed by the thermal bath and the persistent collisions from the active particles, respectively. In the present system $\mathbf{V}_0$ and $\mathbf{V}_A$ are uncorrelated, and $\mathbf{V}_A \ll \mathbf{V}_0$. The active contribution to the mean kinetic energy, $E_A$, can be roughly estimated through Eq. (4) as, $E_A \sim \frac{1}{2} M \langle \mathbf{V}_A^2 \rangle \sim \frac{1}{2} \tau \int_0^\tau \dot{\mathbf{r}}^2 dt = \frac{M \langle \dot{\mathbf{r}}^2 \rangle}{2\tau} [1 - \exp(-2\tau k/\gamma)]$, which monotonically decreases with $k$ and saturates at large $k$, consistent with the simulation measurement in Fig. 4(a) and (c).

Figures 4(b) and 4(d) show that the theoretical values of $U$ and $\tau$ are uncorrelated, and $\tau$ decreases with $k$, which further confirms that the active noise and effective temperature of the confined passive tracer in active baths are not intrinsic properties of active baths, and great care must be taken when applying these quantities in active systems. Our findings are experimentally verifiable by optically trapping a colloidal sphere in a bacterial solution.

**CONCLUSION**

Active noise provides us an ideal framework to coarse grain active bath. Using computer simulations and theoretical calculations, we here show that the active noise experienced by a harmonically trapped passive tracer in an active bath, as well as three related effective temperature, depend sensitively on the constraint imposed on the passive particle, fundamentally different from the equilibrium cases. This constraint dependence arises from the persistent collisions between the passive tracer and the active swimmers, which can be greatly impacted by the constraint strength. Our results unequivocally demonstrate that the active noise and effective temperatures are not intrinsic properties of active baths, and great care must be taken when applying these quantities in active systems. Our findings are experimentally verifiable by optically trapping a colloidal sphere in a bacterial solution.

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